

101.215-12 – including *Taq* pol., IFU-01
101.215-12u – without *Taq* pol., IFU-02

Visit www.olerup-ssp.com for
“Instructions for Use” (IFU)

Lot No.: **47V**

Lot-specific Information
Olerup SSP® DQB1*04

Product number:	101.215-12 – including <i>Taq</i> polymerase 101.215-12u – without <i>Taq</i> polymerase
Lot number:	47V
Expiry date:	2016-September-01
Number of tests:	12
Number of wells per test:	14+1
Storage - pre-aliquoted primers:	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

This Product Description is only valid for Lot No. 47V.

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

**CHANGES COMPARED TO THE PREVIOUS OLERUP SSP®
DQB1*04 LOT (62R)**

A well containing Negative Control primer pairs has been added.

The format of the Product Insert and Worksheet have been changed.

Seven wells have been added to DQB1*04, wells **9 to 15**.

The DQB1*04 specificity and interpretation tables have been updated with the DQB1 alleles described since the previous *Olerup SSP®* DQB1*04 lot (Lot No. 62R) was made.

As of lot series V, the Specificity Table is included in the lot-specific Product Insert, and the Interpretation Table is included in the Worksheet.

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The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
2	-	-	Strength of control band has been optimized.
3	-	-	Strength of control band has been optimized.
5	Added	-	5'-primer added from well 6.
6	Moved, added	Moved, added	Primer pair moved to well 5, primer pair added for the DQB1*04:16 allele.
9	New	New	New primer pair for the DQB1*04:09 and DQB1*04:14 alleles.
10	New	New	New primer pair for the DQB1*04:10 allele.
11	New	New	New primer pair for the DQB1*04:11 and DQB1*04:15 alleles.
12	New	New	New primer pair for the DQB1*04:18 allele.
13	New	New	New primer pair for the DQB1*04:13 allele.
14	New	New	New primer pair for the DQB1*04:17 allele.
15	New	New	Negative Control.

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Lot-specific Information

Well 15 contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup SSP®* HLA Class I, DRB, DQB1 and DPB1 amplicons as well as amplicons generated by a control primer pair.

PCR product sizes range from 75 to 430 base pairs.
The PCR product generated by the control primer pair is 430 base pairs.

Length of PCR product	105	200	105	80	75	80
5'-primer¹	164	340	440	45	45	43
	5'-CAC ^{3'}	5'-Agg ^{3'}	5'-TTA ^{3'}	5'-Tgg ^{3'}	5'-Tgg ^{3'}	5'-Tgg ^{3'}
3'-primer²	231	2nd I	507	59	58	57
	5'-TgC ^{3'}	5'-AAA ^{3'}	5'-TTg ^{3'}	5'-CTC ^{3'}	5'-ggC ^{3'}	5'-CTC ^{3'}
A*	+	+	+			
B*	+	+	+			
C*	+	+	+			
DRB1				+	+	
DRB3				+	+	
DRB5				+		
DQB1					+	
DPB1						+

¹The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

²The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon or the 2nd intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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Lot-specific Information
PRODUCT DESCRIPTION

DQB1*04 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the DQB1*04:01 to DQB1*04:18 alleles.

Please note that DQB1 amplifications usually are somewhat less pronounced than e.g. DRB and DQA1 amplifications even when using the same DNA preparation and exactly the same experimental procedures.

PLATE LAYOUT

Each test consists of 15 PCR reactions in a 16 well PCR plate.

Well 16 is empty

1	2	3	4	5	6	7	8
9	10	11	12	13	14	NC	empty

The 16 well PCR plate is marked with 'DQ4' in silver/gray ink.

Well No. 1 is marked with the Lot No. '47V'.

Wells 1 to 14 – DQB1*04 high resolution primers.

Well 15 – Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.

The PCR plates are covered with a PCR-compatible foil.

Please note: When removing each 16 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Due to the sharing of sequence motifs between DQB1 alleles non-DQB1*04 alleles will be amplified by primer mixes 4, 5, 8 and 10.

Thus, the interpretation of DQB1*04 subtypings is not influenced by other groups of the DQB1 alleles or the DQB2 and DQB3 genes.

For further details see Specificity Table.

UNIQUELY IDENTIFIED ALLELES

All the DQB1*04 alleles, i.e. **DQB1*04:01 to DQB1*04:18**, recognized by the HLA Nomenclature Committee in January 2014^{1,2} will give rise to unique amplification patterns by the primers in the DQB1*04 subtyping kit.

The following DQB1*04 alleles can be distinguished by the different sizes of the specific PCR product:

Alleles	Primer mix
DQB1*04:04, 04:12	5
DQB1*04:05, 04:06	5

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The DQB1*04 subtyping kit cannot distinguish the silent mutations in the DQB1*04:01:01-04:01:04 alleles or the DQB1*04:02:01-04:02:05 alleles.

¹HLA-DQB1 alleles listed on the IMGT/HLA web page 2014-January-17, release 3.15.0, www.ebi.ac.uk/imgt/hla.

²Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page <http://hla.alleles.org/alleles/deleted.html>.

RESOLUTION IN HOMO- AND HETEROZYGOTES

Results file with resolution in DQB1*04 homo- and heterozygotes is available upon request.

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Lot-specific Information
SPECIFICITY TABLE

DQB1*04 SSP subtyping

Specificities and sizes of the PCR products of the 14+1 primer mixes used for DQB1*04 SSP subtyping

Primer Mix	Size of spec. PCR product ¹	Size of control band ²	Amplified DQB1*04 alleles ³	Amplified non-DQB1*04 alleles ⁴
1 ⁶	210 bp	515 bp	*04:01:01-04:02:01, 04:02:03-04:03:01, 04:04-04:18	
	245 bp		*04:01:01-04:03:02, 04:06-04:18	
2 ⁶	205 bp	515 bp	*04:01:01-04:01:04, 04:05-04:08, 04:14-04:17	
3	205 bp	430 bp	*04:02:01-04:03:01, 04:04, 04:09-04:13, 04:18	
4	195 bp	430 bp	*04:03:01-04:03:02	*03:06, 03:25
5 ⁵	110 bp	430 bp	*04:06, 04:12	*03:06, 03:25
	245 bp		*04:04-04:05	
6 ⁵	95 bp	430 bp	*04:16	
7	160 bp	430 bp	*04:07	
8 ⁵	95 bp	430 bp	*04:08	*03:06 [?] -03:08 [?] , 03:10:02 [?] -03:15 [?] , 03:17:01 [?] -03:18 [?] , 03:19, 03:20 [?] , 03:23 [?] , 03:26 [?] , 03:37 [?] , 03:40 [?] , 03:48 [?] , 03:52 [?] -03:71 [?] , 03:74 [?] -03:78 [?] , 03:81 [?] -03:82 [?] , 03:101 [?] -03:112 [?]
9	140 bp	430 bp	*04:09, 04:14	
10	145 bp	430 bp	*04:10	*06:03:03 ^w
11 ⁵	120 bp	430 bp	*04:11, 04:15	
12	230 bp	430 bp	*04:18	
13	185 bp	430 bp	*04:13	
14	160 bp	430 bp	*04:17	
15 ⁷			Negative Control	

¹ Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DQB1*04 SSP typings.

When the primers in a primer mix can give rise to HLA-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

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PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

³For several DQB1 alleles 1st and/or 3rd exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

⁴Due to the sharing of sequence motifs between DQB1 alleles non-DQB1*04 alleles will be amplified by primer mixes 4, 5, 8 and 10.

⁵HLA-specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

⁶Primer mixes 1 and 2 may give rise to a lower yield of HLA-specific PCR product than the other DQB1*04 primer mixes.

⁷Primer mix 15 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by control primer pairs. PCR product sizes range from 75 to 200 base pairs. The PCR product generated by the control primer pair is 430 base pairs.

'w', might be weakly amplified.

'?', nucleotide sequence information not available for the primer matching sequence.

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PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10	11	12
Length of spec. PCR product	210	205	205	195	110	95	160	95	140	145	120	230
Length of int. pos. control ¹	515	515	430	430	430	430	430	430	430	430	430	430
5'-primer(s) ²	9(122) 5'-gTT 3'	23(164) 5'-gCT 3'	23(164) 5'-gCg 3'	26(173) 5'-TCT 3'	9(122) 5'-gTA 3'	59(272) 5'-CgT 3'	144(529) 5'-CCg 3'	167(596) 5'-gCA 3'	43(226) 5'-ACA 3'	23(164) 5'-gCg 3'	48(241) 5'-ggA 3'	15(140) 5'-gTA 3'
	21(159) 5'-ACC 3'		23(164) 5'-gCg 3'		54(259) 5'-ggT 3'				45(230) 5'-ggA 3'		52(251) 5'-gCT 3'	
3'-primer(s) ³	77(327) 5'-ACg 3'	77(327) 5'-ACg 3'	77(327) 5'-ACg 3'	77(327) 5'-ACg 3'	77(327) 5'-ACg 3'	77(327) 5'-ACg 3'	185(650) 5'-CgA 3'	185(650) 5'-CgA 3'	77(327) 5'-ACg 3'	57(267) 5'-gCA 3'	77(327) 5'-ACg 3'	77(327) 5'-ACg 3'
Well No.	1	2	3	4	5	6	7	8	9	10	11	12

Well No.	13	14
Length of spec. PCR product	185	160
Length of int. pos. control ¹	430	430
5'-primer(s) ²	139(514) 5'-CAA 3'	146(533) 5'-CCT 3'
3'-primer(s) ³	187(656) 5'-ACA 3'	185(650) 5'-CgA 3'
Well No.	13	14

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

²The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

³The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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Lot-specific Information

CELL LINE VALIDATION SHEET																		
DQB1*04 SSP subtyping kit ²																		
				Production No.	Well													
					1	2	3	4	5	6	7	8	9	10	11	12	13	14
					201432101	201432102	201432103	201432104	201432105	201432106	201432107	201432108	201432109	201432110	201432111	201432112	201432113	201432114
	IHWC cell line ¹		DQB1															
1	9001 SA		*05:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
2	9280 LK707		*06:01	*02:02	-	-	-	-	-	-	-	-	-	-	-	-	-	-
3	9011 E4181324		*06:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
4	9275 GU373		*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
5	9009 KAS011		*05:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
6	9353 SM		*03:02	*06:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
7	9020 QBL		*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
8	9025 DEU		*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
9	9026 YAR		*03:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
10	9107 LKT3		*04:01		+	+	-	-	-	-	-	-	-	-	-	-	-	-
11	9051 PITOUT		*02:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
12	9052 DBB		*03:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-
13	9004 JESTHOM		*05:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
14	9071 OLGA		*04:02		+	-	+	-	-	-	-	-	-	-	-	-	-	-
15	9075 DKB		*03:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-
16	9037 SWEIG007		*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
17	9282 CTM3953540		*02:01	*06:03	-	-	-	-	-	-	-	-	-	-	-	-	-	-
18	9257 32367		*06:02	*02:02	-	-	-	-	-	-	-	-	-	-	-	-	-	-
19	9038 BM16		*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
20	9059 SLE005		*06:04		-	-	-	-	-	-	-	-	-	-	-	-	-	-
21	9064 AMALA		*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
22	9056 KOSE		*05:03	*06:04	-	-	-	-	-	-	-	-	-	-	-	-	-	-
23	9124 IHL		*05:03	*06:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
24	9035 JBUSH		*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
25	9049 IBW9		*02:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
26	9285 WT49		*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
27	9191 CH1007		*04:01	*05:01	+	+	-	-	-	-	-	-	-	-	-	-	-	-
28	9320 BEL5GB		*02:02	*03:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
29	9050 MOU		*02:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
30	9021 RSH		*04:02		+	-	+	-	-	-	-	-	-	-	-	-	-	-
31	9019 DUCAF		*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
32	9297 HAG		*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
33	9098 MT14B		*03:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
34	9104 DHIF		*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
35	9302 SSTO		*03:05		-	-	-	-	-	-	-	-	-	-	-	-	-	-
36	9024 KT17		*03:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
37	9065 HHKB		*06:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-
38	9099 LZL		*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
39	9315 CML		*02:01	*03:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
40	9134 WHONP199		*02:02	*03:03	-	-	-	-	-	-	-	-	-	-	-	-	-	-
41	9055 H0301		*06:09		-	-	-	-	-	-	-	-	-	-	-	-	-	-
42	9066 TAB089		*06:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
43	9076 T7526		*03:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-
44	9057 TEM		*05:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-
45	9239 SHJO		*02:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
46	9013 SCHU		*06:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-
47	9045 TUBO		*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-
48	9303 TER-ND		*05:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-

¹The provided cell line HLA specificities are retrieved from the <http://www.ihwg.org/hla> web site. The specificity of an individual cell line may thus be subject to change.

²The specificity of each primer solution in the kit has been tested against 48 well characterized cell line



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DNAs and where applicable, additional cell line DNAs

No DNAs carrying the alleles to be amplified by primer solutions 4 to 7 and 9 to 14 were available. The specificities of the primers in primer solutions 4, 5 and 9 were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer. In primer solutions 6, 7, 11, 12 and 14 it was only possible to test the 3'-primer, the 5'-primers were not possible to test. In primer solutions 10 and 13 it was only possible to test the 5'-primer, the 3'-primer was not possible to test. In primer solutions 3, 5 and 9 one 5'-primer was not possible to test.

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Lot-specific Information

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